

Telephone: 814.272.1039 Fax: 814.272.1019

# **Analytical Report**

# PFOA and PFOS Analysis of Wild Turkey Muscle and Liver Samples by LC/MS/MS

MPI Report No. L0019897

## **Testing Laboratory**

MPI Research, Inc. 3058 Research Drive State College, PA 16801

# Requester/Project Manager

Dena Haverland Dalton Utilities PO BOX 869 Dalton, GA 30722 Phone: 706-529-1010

#### Introduction

Results are reported for the analysis of turkey muscle and liver samples received at MPI Research from Dalton Utilities. The MPI Research study number assigned to the project is L0019897. Table I lists the target analytes quantitated for the samples.

Table I. Target Analytes for Quantitation

Compound Name	Acronym
Perfluorooctanoic Acid	C8 Acid or PFOA
Perfluorooctanesulfonate	C8 Sulfonate or PFOS

#### 2 Sample Receipt

Four samples were received from Dena Haverland at Dalton Utilities for this study. The samples were collected on December 16, 2009. The samples arrived on December 22, 2009 via Fedex and were logged in under MPI Research login number L0019897. The shipment was received frozen on dry ice. The samples were stored frozen at approximately -20°C from receipt until analysis. Chain-of-custody information is presented in Attachment A.

## 3 Methods - Analytical and Preparatory

## 3.1 Muscle and Liver Sample Preparation

- 3.1.1. Weigh 1 g of muscle or liver sample into a 50 mL disposable centrifuge tube and fortify, if appropriate. Add appropriate amount of internal standard solution.
- 3.1.2 Add water to the sample for a final volume of 10 mL. Cap tightly.
- 3.1.3 Homogenize sample using a tissuemizer for ~1 minute.
- 3.1.4 Transfer 1 mL of the sample using a disposable pipette into 15 mL disposable centrifuge tubes. Add 5 mL of ACN and shake for ~20 minutes on a wrist action shaker.
- 3.1.5. Centrifuge tubes at ~3000 rpm for ~ 5 minutes. Carefully decant supernatant into a 50 mL disposable centrifuge tube and add 35 mL of water.
- 3.1.6 Place the unconditioned SPE columns on the vacuum manifold. Condition the SPE columns by passing ~ 10 mL of methanol through the column followed by ~ 5 mL of water. The washes may be pulled through the SPE column using vacuum at a flow rate of ~1 drop/sec or may be allowed to pass through the column unaided. Discard all washes. Do not allow the column to dry.
- 3.1.7 Load the sample onto a conditioned SPE column. Discard the eluate. Any analyte residues will be trapped on the SPE column at this point.
- 3.1.8 Elute with 2 mL of methanol. Collect 2 mL of elute into a graduated 15 mL centrifuge tube.

Note: Post extraction dilutions were prepared in methanol.

#### 3.2 Sample Analysis by LC/MS/MS

In High Pressure Liquid Chromatography (HPLC), an aliquot of extract is injected and passed through a liquid-phase chromatographic column. Based on the affinity of the analyte for the stationary phase in the column relative to the liquid mobile phase, the analyte is retained for a characteristic amount of time. Following HPLC separation, mass spectrometry provides a rapid and accurate means for analyzing a wide range of organic compounds. Molecules are ionized, fragmented, and detected. The ions characteristic of the compounds are observed and quantitated against external calibration standards.

An HP1100 system interfaced to an Applied Biosystems API 4000 LC/MS/MS was used to analyze the sample extracts for quantitation. A gradient elution through a Phenomenex Luna  $3\mu$  C8(2) Mercury, 20 x 4.0 mm column was used for separation.

The following gradient was performed:

2mM Ammonium Acetate in Water Methanol				

The following parameters were used for operation of the mass spectrometer:

Parameter	Setting
Ionization Mode	Electrospray
Polarity	Negative
Transitions Monitored	413→369 (PFOA)
	413→219 (PFOA Confirmation)
	499→80 (PFOS)
	499→99 (PFOS Confirmation)
	415→370 (Internal Std. <sup>13</sup> C PFOA (m+2))
	503→80 (Internal Std. <sup>13</sup> C PFOS (m+4))
Gas Temperature	450°C

## 4 Analysis by LCMSMS

#### 4.1 Calibration

For the muscle and liver sample analysis, a 6-point calibration curve was analyzed throughout the analytical sequence for PFOS. The calibration points were prepared at 0.1, 0.2, 0.5, 1.0, 2.0, 5.0 ng/mL (ppb) containing 1.0 ng/mL <sup>13</sup>C-PFOS (m+4).

The ratio of the analyte concentration to the IS concentration versus the ratio of the analyte instrument response (area) to the IS response (area) was plotted for each point. Using linear regression with 1/x weighting, the slope, y-intercept and coefficient of determination ( $r^2$ ) were determined. A calibration curve is acceptable if  $r^2 \ge 0.985$ .

For the results reported here, calibration criteria were met. The calibration curves are included in the raw data in Attachment C.

#### 4.2 Laboratory Control Spikes

Laboratory control spikes in the analytical set were prepared during each extraction set by adding a known concentration of the analyte to turkey muscle and liver controls. Laboratory control spikes are used to assess method accuracy. The laboratory control spikes must show recoveries between 70-130% or the data is rejected. For the results reported here, the laboratory control spikes were within the acceptable range. Laboratory control spike recoveries are given in Attachment B.

#### 4.3 Matrix Spikes

A matrix spike was prepared for each sample by adding a known concentration of the target analyte to a sample. Matrix spikes are used to assess method accuracy in the matrix. The matrix spikes should show recoveries between 70-130%. For the results reported here, the matrix spike was within the acceptable range with the exceptions of:

#### 4.4 Laboratory Duplicates

Each sample was prepared in duplicate and analyzed. Duplicate results are given along with the sample results in Attachment B.

#### 5 Data Summary

Due to an interfering matrix peak at the 499→80 m/z transition, the 499→99 m/z confirmation transition was used for quantitation. The interfering matrix peak was not present at the 499→99 m/z confirmation transition.

Please see Attachment B for a detailed listing of the analytical results. For the muscle and liver samples the results are reported in parts per billion (ng/g) on an as-received basis.

## 6 Data/Sample Retention

Samples are disposed of 60 days after the report is issued unless otherwise specified by the project manager. All electronic data is archived on retrievable media and hard copy reports are stored in data folders maintained by MPI Research. Hardcopy data is stored for a minimum of five years. The client will be notified 30 days prior to the disposal of hardcopy data.

## 7 Attachments

- 7.1 Attachment A: Chain of Custody
- 7.2 Attachment B: Analytical Results
- 7.3 Attachment C: Raw Analytical Data

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Mark Neeley, Research Chemist Associate II Date

Robert Zhu, Manager, Analytical Date



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Mattawan (Corporate Headquarters)

Conform COC Sample:

Conform COC:

Conform Sample:

Conform Request:

54943 North Main Street Mattawan, MI 49071-9399 (269) 668-3336 Phone (269) 668-4151 Fax

State College 3058 Research Drive State College, PA 16801 (814) 272-1039 Phone (814) 231-1580 Fax

True

True

True

True

Login

Login Group: L0019897

Login #:

20011

Project:

P0005380 **Dalton Utilities** 

Company Name: Submitted By:

Dena Haverland

Login Type:

Immediate Receipt of Samples

Started: Date Start: True 12/31/2009

Due Date: Login Initiated:

01/10/2010 12/31/2009

Kyle, Matt

Received By:

Spread Sample:

Label:

2/3/2010

Login.rpt

MPI SD/PI: Zhu, Xiang

Project Title/Type: PFOA AND PFOS ANALYSIS OF TURKEY BLOOD, MUSCLE AND LIVER / ROUTINE

Login Notes:

•		<u>Packag</u>	es / Containers		
<u>Package</u>	Carton	Date / Condition	Shipper / ID	Temp. Control/Temp.	Direction / Handled By
PK0022626	Received Package	d Date: 12/22/09 11:33 & Contents Uncompromised	FEDEX 8694 2057 8384	Dry Ice -79.0	RECEIVED Kyle, Matt
Container # 3182	Gross Weight 417.70 g	pH <u>Container Type</u> 1 gallon ziploc bag	Preservative NONE	Mfg. Lot	Mfg. ID
C0473183	143.40 g	1/2 gallon ziplock bag	NONE		
C0473184	458.00 g	1 gallon ziploc bag	NONE		
C0473185	89.90 g	1/2 gallon ziplock bag	NONE		

				Samples	B		
<u>Sample ID</u> L0019897-0001	Container C0473182	<u>Matrix</u> SOLID	<u>System</u> Deer	System Matrix Tissue	Sample Wild Turkey #2 4yr Male - Muscle	Date Sampled 12/16/2009	<u>Date Due</u> 01/10/2010
L0019897-0002	C0473183	SOLID	Deer	Liver	Wild Turkey #2 4yr Male - Liver	12/16/2009	01/10/2010
L0019897-0003	C0473184	SOLID	Deer	Tissue	Wild Turkey #5 1yr Male - Muscle	12/16/2009	01/10/2010
L0019897-0004	C0473185	SOLID	Deer	Liver	Wild Turkey #5 1yr Male - Liver	12/16/2009	01/10/2010



Instance:

Report Version: Jan 7 2009 1:39PM

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MPI Research Contact: \_\_\_\_\_Daniel Wright

# 

# Sample Submittal

Please fax this form before sending samples.

Please send samples to shipping and receiving: 3048 Research Drive, State College, PA 16801 T: (814) 272-1039 • F: (814) 272-1019					
Turnaround time (TAT) requirements:					
Results Due Date: 30 days					
Preliminary Results Format: Verbal Email Fax					
Report Due Date: 3	odays				
Storage Conditions	Safety Information				
Room temperature	Special handling:				
Refrigerator Freezer	MSDS attached				
Ultra Low freezer Desiccated	Controlled substance:				
Lighting required	HAZARDS:				
Stability (°C/%RH):	Please fill in the diamond HMIS/NFPA				

	Client ID# Description	Lot/ Control #	Amt. Sent/ Weight	# of Bottles	Matrix	Date & Time	Tests Requested
1	Wild Turkey #2 4yr Male - Scrum	1000	3m1	2	Turkey	12/16/69 11:15 am	PFOA/PFOS
2	Wild Turkey #2 4vr Male - Muscle					12/16/09 11:15 am	
3	Wild Turkey #2					12/16/09 11:15 am	
4	Wild Turkey #5 Tyr Male - Serum					12/16/09 12:00 pm	
5	Wild Turkey #5					12/16/09 12:00 pm	
6	Wild Turkey #5 lyr Male-Liver			i	1	12/16/09 12:00 pm	1
7					7		
8							
9							
10							

Stability time period:

Relinquished by	Date	Time	Received by	Date	Time
A. 111	112/21/10	3:00pm	111	12/2/2	//93
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FedEx USAirbill Tends B694 2057 B384	1 From  Date 12/21/09  Sender's Darrell Karrarattap Phone 206 546-5627	Address 2 C. P. J. C. A. C. A. C. A. C. A. C. A. C.	2 Your Internal Billing Reference 3 To Recipient's	MPI Receipt habe	Address Address To request a package be held at a specific Fadex location, print Fadex address hare.	8894 2057 8384

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THE ORIGINAL DOCUMENT"

BY ME DATE 12/2/1



## **TEMPORARY SAMPLE STORAGE FORM**

To be completed during ExyLIMS Login
Project #: <b>Ps194</b>
Login #:
Initials / Date:
One form to be completed for each package
Date / Time Received: אַכּוֹנוֹ פּאַרוּ אַפּאַנוּ אַפּאַנוּ אַפּאַנוּ אַפּאַנוּ אַפּאַנוּ אַפּאַנוּ אַפּאַנוּ
Received By: M.K.
Shipper: FLL EX
Shipper Package ID: 8694 2017 8384
Temperature (deg C) / Thermometer ID:
Temperature Control Method: Dry lu
Temporary Storage Location: Walky Freeze 1
Condition of sample(s): Good – Package and contents uncompromisedFair – Package damaged / contents uncompromisedPoor – Package and contents compromised
Notes:

B



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# **Analytical Report**

# **Summary of Fluorochemical Residues in Muscle Samples**

	PFOA	PFOS		
	Perfluorooctanoic Acid	Perfluorooctanesulfonate		
	Analyte	Analyte		
	Found	Found		
Sample ID	(ng/g, ppb)	(ng/g, ppb)		
Wild Turkey # 2 4yr male-muscle	ND	82.6		
Wild Turkey # 2 4yr male-muscle*	NQ	89.6		
Wild Turkey # 5 1yr male-muscle	NQ	176		
Wild Turkey # 5 1yr male-muscle*	NQ	193		

<sup>\*</sup>Laboratory Duplicate

ND = Not detected = Response is below the LOD of 1.0 ng/g (ppb).

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g (ppb).



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# **Analytical Report**

# **Summary of Fluorochemical Residues in Liver Samples**

	PFOA Perfluorooctanoic Acid	PFOS Perfluorooctanesulfonate		
Sample ID	Analyte Found (ng/g, ppb)	Analyte Found (ng/g, ppb)		
Wild Turkey # 2 4yr male-liver	NQ	1560		
Wild Turkey # 2 4yr male-liver*	NQ	1580		
Wild Turkey # 5 1yr male-liver	12.8	2140		
Wild Turkey # 5 1yr male-liver*	12.3	1970		

<sup>\*</sup>Laboratory Duplicate

ND = Not detected = Response is below the LOD of 1.0 ng/g (ppb).

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g (ppb).



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# Recovery Summary of Fluorochemical Residues in Muscle Samples

			PFOA			PFOS	
Sample Description	Amount Spiked (ng/g	Amt Found in Sample (ng/g)	Amount Recovered (ng/g)	Recovery (%)	Amt Found in Sample (ng/g)	Amount Recovered (ng/g)	Recovery (%)
LCS A (Data set 020910A) 10 ng/g	10	NQ	9.77	98	ND	8.05	81
LCS B (Data set 020910A) 50 ng/g	50	NQ	49.6	99	ND	54.3	109
LCS A (Data set 021210A) 200 ng/g	200	N/A	N/A	N/A	ND	205	103
LCS B (Data set 021210A) 200 ng/g	200	N/A	N/A	N/A	ND	171	86
Wild Turkey # 2 4yr male-muscle (L19897-1 Spk C, 50 ng/g Lab Spike)	50	ND	49.5	99	N/A	N/A	N/A
Wild Turkey # 5 1yr male-muscle (L19897-3 Spk D, 50 ng/g Lab Spike)	50	ND	50.6	101	176	212	72
Wild Turkey # 2 4yr male-muscle (L19897-1 Spk C, 200 ng/g Lab Spike)	200	N/A	N/A	N/A	82.6	238	78

ND = Not detected = Response is below the LOD of 1.0 ng/g.

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g.



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## **Recovery Summary of Fluorochemical Residues in Liver Samples**

				PFOA			PFOS	
	Sample Description	Amount Spiked (ng/g	Amt Found in Sample (ng/g)	Amount Recovered (ng/g	Recovery (%)	Amt Found in Sample (ng/g	Amount Recovered (ng/g	Recovery (%)
	LCS A (Data set 020910B) 10 ng/g	10	ND	10.3	103	ND	7.64	76
	LCS B (Data set 020910B) 50 ng/g	50	ND	49.6	99	ND	65.8	132
	LCS A (Data set 021210B) 2000 ng/g	2000	N/A	N/A	N/A	ND	1880	94
:	LCS B (Data set 021210B) 2000 ng/g	2000	N/A	N/A	N/A	ND	2080	104
	Wild Turkey # 2 4yr male-liver (L19897-2 Spk C, 50 ng/g Lab Spike)	50	NQ	58.4	117	1560	**	**
	Wild Turkey # 5 1yr male-liver (L19897-4 Spk D, 50 ng/g Lab Spike)	50	12.8	63.3	101	2140	**	**
	Wild Turkey # 2 4yr male-liver (L19897-2 Spk C, 2000 ng/g Lab Spike)	2000	N/A	N/A	N/A	1560	3870	116
	Wild Turkey # 5 1yr male-liver (L19897-4 Spk D, 2000 ng/g Lab Spike)	2000	N/A	N/A	N/A	2140	4110	99

ND = Not detected = Response is below the LOD of 1.0 ng/g.

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g.